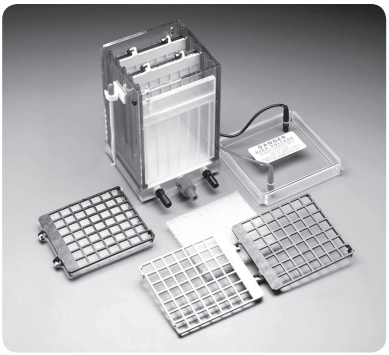
Hoefer™ TE 22

tank transfer unit



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Hoefer

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English

Safety warnings and precautions

Important user information

Please read this entire manual to fully understand the safe and effective use of this product.

The exclamation mark within an equilateral triangle is intended to alert the user to the presence of important operating and maintenance instructions in the literature accompanying the instrument.

The lightning symbol within an equilateral triangle is intended to alert the user to the risk of exposure to high voltages.

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Français

Renseignements importants d'utilization

Pour une bonne compréhension et une utilisation en sécurité maximale, il convient de lire entièrement ce manuel.

Dans la documentation qui accompagne l'instrument un point d'exclamation dans un triangle équilatéral a pour but d'attirer l'attention de l'utilisateur sur des instructions importantes de fonctionnement ou de maintenance.

Le symbole de l'éclair dans un triangle équilatéral a pour objet d'attirer l'attention de l'utilisateur sur un danger d'exposition à la haute tension.

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Deutsch

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Español

Información importante para el usuario

Para comprender el producto y utilizarlo con seguridad es necesario leer este manual en su totalidad.

El signo de admiración en un triángulo equilátero en el manual, advierte al usuario sobre la presencia de instrucciones importantes de operación y mantenimiento del aparato.

El símbolo del rayo en un triángulo equilátero alerta al usuario sobre el riesgo de exposición a altas tensiones.

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Italiano

Informazioni importanti per l'operatore

Per un utilizzo sicuro del prodotto, leggere attentamente l'intero contenuto del presente manuale.

Il punto esclamativo all'interno di un triangolo equilatero indica all'operatore la presenza di importanti istruzioni di funzionamento e manutenzione nella documentazione allegata al prodotto.

Il simbolo del fulmine all'interno di un triangolo equilatero indica all'utente la presenza di un rischio di esposizione ad alte tensioni.

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Transfer Electrophoresis Unit function and description

The Hoefer™ TE22 Tank Transfer unit rapidly transfers proteins, DNA, or RNA from up to four small-format polyacrylamide or agarose gels onto a membrane. Gels and membranes are held by a cassette, which is submerged into the transfer tank. Molecules migrate under an electric field to the membrane, where they are bound.

The transfer buffer temperature can be controlled by circulating cooled liquid through the heat exchanger in the base. The buffer is separated from the coolant by a heat-conducting alumina plate.

Unpacking

Unwrap all packages carefully and compare contents with the packing list, making sure all items arrived. If any part is missing, contact Hoefer, Inc.. Inspect all components for damage that may have occurred while the unit was in transit. If any part appears damaged, contact the carrier immediately. Be sure to keep all packing material for damage claims or to use should it become necessary to return the unit.

This declaration of conformity is only valid for the instrument when it is:

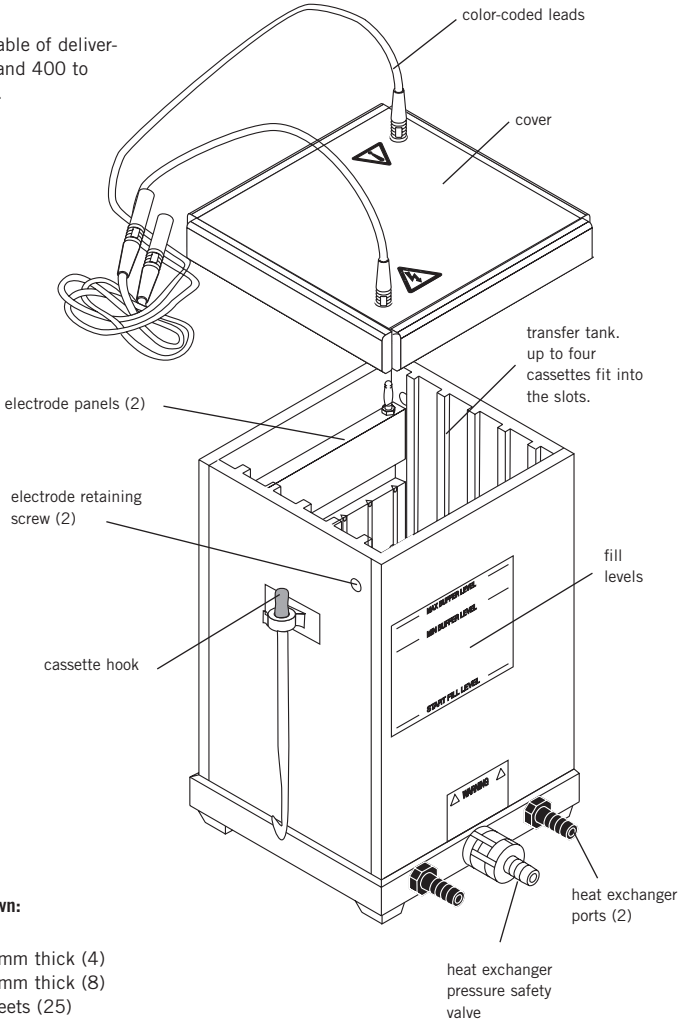
- used in laboratory locations,
- used as delivered from Hoefer, Inc. except for alterations described in the User Manual, and
- connected to other CE labeled instruments or products recommended or approved by Hoefer, Inc.

Specifications

Gel size	up to four 9 × 10 cm gels
Max. wattage	50 W
Max. voltage	100 V≂
Max. amperage	500 mA
Max. temperature	45 °C
Buffer required	1.5 liters, depending on the number of cassettes in place
Environmental operating conditions	Indoor use: 4–40 °C Humidity up to 80% Altitude up to 2000 m
Installation category	II
Pollution degree	2
Dimensions (w × h × d)	14 × 24 × 16.5 cm (5.5 × 9.5 × 6.5 in)
Product certifications	EN61010–1, UL3101–1, CSA C22.2 1010.1, CE

Fig 1. Tank transfer unit main components.

A power supply capable of delivering up to 100 V≂ and 400 to 500 mA is required.



Included but not shown:

- Gel cassettes (4)
- Foam sponges, 6 mm thick (4)
- Foam sponges, 3 mm thick (8)
- Blotting paper, sheets (25)

English

Important information

- The safety lid must be in place before connecting the power leads to a power supply.
- Turn all power supply controls off and disconnect the power leads before removing the safety lid.
- Rinse only the electrodes (not the banana plugs) with distilled water before use.
- Circulate only water or 50/50 water/ethylene glycol through the heat exchanger. Never use anti-freeze mixtures in the heat exchanger.
- **Do not operate with buffer temperature above 45 °C.** All plastic parts are rated for 45 °C continuous duty.
- Circulate coolant through the heat exchanger to minimize heating. Overheating will cause irreparable damage to the unit! Do not connect the heat exchanger to a water tap or any coolant source where the water pressure is unregulated.
- For longer runs you can control heating somewhat by chilling the buffer before use, running the unit in a cold room, or both.
- When assembling the transfer cassette, use only the required amount of gel support materials (sponges and blotting paper) to prevent overstuffing the cassette. **Excess materials may result in cassette damage.**
- Never expose any part of the instrument to alcohols or organic solvents.* Alcohols or organic solvents will cause irreparable damage to the unit!
- If this equipment is used in a manner not specified by the manufacturer, the protection provided by the equipment may be impaired.
- Only accessories and parts approved or supplied by Hoefer, Inc. may be used for operating, maintaining, and servicing this product.

*Use of ≤20% methanol (methyl alcohol) in transfer buffers is the only exception.

Français

Informations importantes

- Le couvercle de sécurité doit être en place avant de brancher les prises au générateur.
- Eteindre le générateur et débrancher les prises avant d'enlever le couvercle de sécurité.
- Rinser seulement les électrodes (pas les "banana-plugs") avec de l'eau distillée juste avant l'utilisation.
- Faire circuler seulement de l'eau ou bien une solution eau/ethylène glycol dans une proportion de 50:50, dans l'échangeur de chaleur. Ne jamais faire circuler une solution d'antigel dans l'échangeur de chaleur.
- **Ne pas utiliser avec un tampon à une température au dessus de 45 °C.** Toutes les piéces en plastique sont prévues pour résister à une température constante de 45 °C.
- Faire circuler l'eau dans l'échangeur vertical pour minimiser l'échauffement afin d'éviter des dommages irréparables à l'instrument. Ne pas connecter l'échangeur vertical à circulation d'eau à un robinet ou quelque source de refroidissement dont la pression n'est pas régulière.
- Pour des coulages plus long, on peut aussi contrôler la température en refroidissant le tampon avant l'utilisation et/ou en utilisant l'instrument dans une chambre froide.
- Utiliser uniquement la quantité prescrite d'éponges et de papier filtre afin que la cassette ne soit pas trop pleine. **Trop de materiels peut endommager la cassette.**
- Ne jamais exposer l'appareil à des alcools et des solvents organiques.* Les alcools et les solvents organiques causent des dommages irréparables à l'appareil.
- Si l'instrument n'est pas utilisé en conformité avec les recommandations du fabriquant, les protections de sécurité qui équipent cet appareil peuvent être rendues inéfficaces.
- Seulement les accessoires et piéces detachées approuvés ou fournis par Hoefer, Inc. sont recommandés pour l'utilisation, l'entretien et réparation de cet appareil.

*L'utilisation de méthanol à une concentration finale de 20% dans les tampons de transfert constitue la seule exception.

Operating instructions

Perform the transfer as soon as possible after electrophoresis to minimize band diffusion. Each step is described below.

Prepare the buffer

Note: Refer to the Electrotransfer Notes section for a discussion of membranes and buffers.

Prepare a minimum of 1.5 liters of the appropriate transfer buffer. Chill the buffer before use if possible.

Prepare the unit

❶

Rinse the transfer tank and cassettes with distilled water.

❷

Note: For quick and easy connections, install Quick-fit coupler fittings with valves in the line.

Active cooling is optional but strongly recommended. If no active cooling will be used, go to step 3.

Note: Connect the heat exchanger to a circulator bath such as the MultiTemp™ III.

Circulate only water or 50/50 water/ethylene glycol to prevent damage to the unit.

The circulator pump must not generate a pressure greater than 0.7 bar (10 psi) above atmospheric pressure.

Set the temperature to 10 °C or higher if circulating only water. If using 50/50 ethylene glycol/water, the temperature can be set lower.

Start the circulator bath at the same time as the transfer.

First attach tubing to the red pressure relief valve between the water inlet and outlet ports and insert the free end into the bath or other container or drain to catch any pressure relief overflow. The relief valve opens if the pressure within the heat exchanger exceeds 10 psi.

Prepare two lengths of 9 mm (3/8") vinyl or silicone tubing. Slide hose clamps (4 total) onto each end of two lengths of tubing. Attach one end of each length of tubing to a heat exchanger port. Attach the free

ends of each length of tubing to the circulator bath ports; one to the inlet and the other to the outlet. Secure the connections with the hose clamps.

❸

Place (**do not drop**) a magnetic stirring bar in the buffer tank. (Dropping objects into the tank may crack the alumina plate.) Set the unit onto a magnetic stirrer and fill transfer buffer to the "Start fill level" line on the front of the tank. (This requires approximately 0.7 liters.)

❹

Set the stirrer to low-medium, which accomplishes buffer circulation without forcing buffer through the cassettes.

> **Note:** Even if no cooling is required for your system, the buffer should be circulated with a stirrer to avoid buffer depletion at the electrodes.

Assemble the transfer cassette

❶

Note: Always wear gloves when handling membranes to avoid getting fingerprints on them.

Pre-wet nitrocellulose or nylon membranes with distilled water. Pre-wet PVDF or other hydrophobic membranes in methanol. Then soak all membrane types in transfer buffer for 2–5 minutes.

❷

Important! Take great care in removing all air bubbles at each step because the presence of air bubbles, especially between the membrane and gel, blocks transfer.

Open the cassette by releasing both latch tabs along the edge opposite the hinges. Place the opened cassette into a tray filled with at least 3 cm of transfer buffer.

❸

Assemble the transfer stack so that molecules will migrate toward the membrane. For negatively charged macromolecules (such as nucleic acids and most proteins), build the stack on the grey half of the cassette (and then later position the lid so that the grey side faces the red lead, or anode, +).

Place one 3 mm-thick foam sponge on the opened submersed cassette and press gently until all air is expelled. Place one sheet of blotting paper on the

sponge, and then place the membrane on the blotting paper. Place the gel—which contains a sample that has been electrophoretically separated and equilibrated (if required) with transfer buffer—on the membrane. Gently roll a glass pipet or test tube over the gel to expel trapped air between the membrane and gel. Cover the gel with a sheet of blotting paper and then place a sponge of the proper thickness (see the diagram below), again pressing gently to expel trapped air.

4

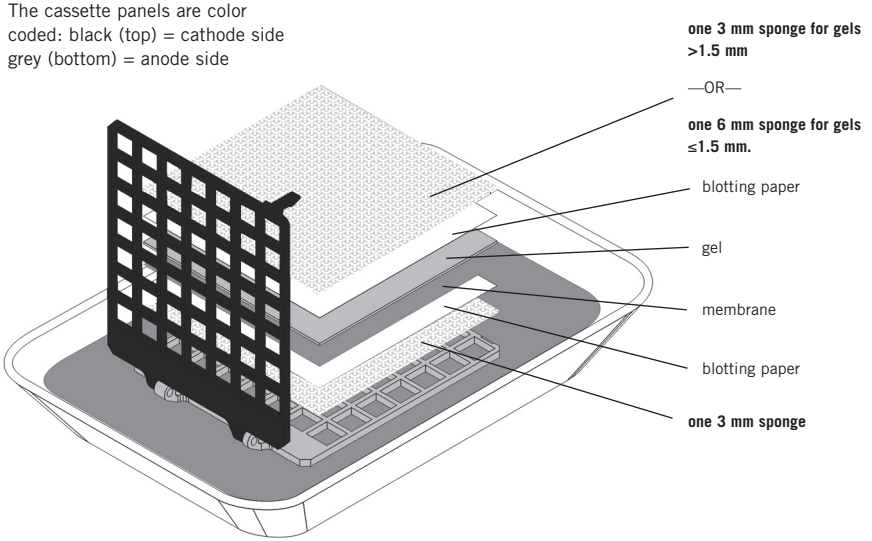
Close the cassette and press lightly to lock the tabs. The assembled cassette should hold the gel in firm contact with the membrane without squeezing the gel. If the stack seems loose, add sheets of blotting paper; if the stack seems tight, replace the top sponge (over the gel) with a sheet of blotting paper. If you remove the bottom sponge (below the gel), substitute at least two sheets of blotting paper to create space between the membrane and the cassette panel.

Fig 2. Transfer stack assembly. The stack is oriented so that negatively charged molecules migrate toward the grey anode, +.

Important! Do not overstuff the cassette.

Note: Try to place the gel correctly the first time because proteins may begin to transfer immediately; once transfer has begun, moving the gel will distort results or cause "shadow bands" on the blot.

The cassette panels are color coded: black (top) = cathode side grey (bottom) = anode side



Assemble the cassette in a tray containing transfer buffer about 3 cm deep.

Install the cassette(s)

❶

If transferring only one or two gels, choose the cassette positions nearest the center. The cassettes must be oriented so that the hinge side is facing up and all black panels of the cassettes are facing the same side of the transfer unit.

Work quickly when moving the assembled cassette(s) to the tank to avoid draining the sponges: Place the tray holding the cassette(s) near the tank, lift out one cassette at a time, and slide it into a set of vertical slots. Do not discard the buffer.

❷

Once in place, tap the cassette lightly until most air bubbles are dislodged. (A few small bubbles in the sponges are unlikely to interfere with the transfer.)

❸

Inspect the buffer level. Add or remove buffer as required so that the level falls between the minimum and maximum buffer level lines. (Buffer above the maximum buffer level line may cause corrosion of the electrical contacts.)

Final assembly and transfer

❶

Install the lid

The cassettes are color coded to match the leads in the lid. To transfer toward the anode, orient the lid so that the grey half of the cassette faces the anode (+), or red lead, and the black half of the cassette faces the cathode (–), or black lead. Make sure the banana plugs seat into the connectors in the lid.

Note: Take care in orienting the lid so that all species migrate toward the membrane when the electric field is applied. The migration direction depends on both the characteristics of the sample and the pH of the transfer buffer. If the species of interest is negatively charged in the transfer buffer and the stack is assembled so that the membrane is nearest the grey side of the cassette, then this side faces the anode (+). Most proteins migrate toward the anode in the Towbin Tris/glycine/methanol buffer system (independent of the presence of SDS), and under most conditions, nucleic acids are negatively charged and also migrate toward the anode.

❷

Use only an approved power supply such as the Hoefer PS 2A200. Make sure the power supply is off and all controls are set to zero. Plug the color-coded leads from the lid of the transfer unit into the power supply—the red lead into the red output jack, and the black lead into the black output jack. In most systems, the red lead is the anode (+), and the black lead is the cathode (–).

❸

Cooling is strongly recommended

Important! Never allow the buffer temperature to exceed 45 °C. Excessive heat will cause the unit to warp.

Any setting that results in higher than 5 W of power will generate enough heat to require active heat control. A refrigerated circulator bath using water should be set to about 10 °C. (If using 50/50 ethylene glycol/water, the temperature can be set lower.) Chill the buffer before use if possible.

Typical transfer parameters

Parameters for your sample and buffer system must be determined empirically.

	protein	nucleic acids
Buffer	Towbin	1X TBE or 1X TAE
Current (A)	0.4	0.3
Voltage (V)	~100	50
Transfer time	~1 hour	~1 hour
Coolant temp.	10 °C	10 °C or less

❹

Set the power supply

Constant current mode is recommended. If constant voltage mode is selected, carefully monitor the current (increased current increases Joule heating). If the current exceeds 0.4 A, decrease the voltage.

❺

If available, set the power supply timer

Most transfers are complete within one hour, but larger molecules or thicker gels may require longer transfer times; the optimum transfer time for each system must be determined empirically.

After the transfer is complete

❶

Turn the voltage and current settings to zero and turn off the power supply. Disconnect the leads from the power supply jacks.

❷

Lift off the lid. Use the plastic hook (stored in the holder at the side of the unit) to lift up a cassette just far enough to be able to grab it and place it into a tray.

❸

Open each cassette carefully and remove the gels and membranes. Label each membrane and indicate the sample side. Lift membrane(s) with blunt forceps and air dry, or follow the instructions accompanying your protocol.

Note: It is a good idea to stain the gel to determine the completeness of the transfer.

Note: Do not store used buffer with transfer tank. Chill buffer to 10 °C before reuse.

❹

Discard the blotting paper, but reuse the sponges.

❺

Rinse the unit immediately after use. (See the Care and maintenance section on the next page.)

Care and maintenance

Cleaning

- Do not autoclave or heat any part above 45 °C.
- Do not expose to alcohols or organic solvents!*
- Never use abrasive detergents.
- If using radioactive reagents, decontaminate the unit with a cleaning agent such as Contrad 70™ or Decon 90™.

Rinse the tank, cassettes, and sponges with distilled water immediately after each use. Allow the unit to air dry completely. Periodically wash with a dilute solution of a mild detergent.

*Use ≤20% methanol (methyl alcohol) in transfer buffers is the only exception.

Removing the electrode panel(s)

For more thorough cleaning or to replace damaged electrodes, remove each electrode panel by unscrewing the retaining screw far enough to allow the panel to slide out. Use the hook on the side panel to pull the electrode panel up (do not pull the panel up by the banana plug). Take care to not stretch or break the platinum wire when handling the panel.

Troubleshooting

problem	solution
Incomplete transfer	
Blank areas on the membrane	Remove all trapped air pockets in the transfer stack assembly: assemble the stack while it is submerged in transfer buffer, gently press on each sponge as it is added to the stack, and roll a glass pipette or test tube over the membrane and gel to eliminate all air bubbles.
	Reduce the stirring speed to prevent turbulence.
	Process only one strip or membrane in each tray or cassette to prevent overlapping.
	Use buffer with a lower ionic strength.
	Check electrode continuity. During the transfer, a continuous stream of gas is released along the entire length of the electrodes. If bubbles do not form along the entire length of the electrode, replace the electrode.
	If cassettes are bowed when empty, replace. Overpacking the cassette causes it to bow; see the recommended assembly instructions on page 7.
Grid pattern on membrane	Add extra sheets of blotting paper to increase the clearance between the cassette panel and the gel. Take care not to overstuff the cassette; the gel should be held firmly and evenly between the sponges, but not so tightly that it is squeezed.
Molecules do not migrate out of gel	Increase the field strength.
	Increase transfer period. (Try doubling it.)
	Do not use staining or fixing agents on the gel before transfer.
	Use a thinner gel.
	Reduce the gel acrylamide concentration.
	Check that the buffer pH is close to the intended pH. Most buffers should not be titrated; make fresh buffer.
	Use 3.5 mM SDS (0.1%) in the transfer buffer.
	Avoid including methanol in the transfer buffer or reduce the amount to the absolute minimum.
	Use reagent-grade chemicals.
	Increase the length of time Southern blots are depurinated.
	Increase the net charge on the protein by changing to a transfer buffer with a different pH. Lower pH (<6–7) increases the positive charge on proteins; higher pH (>6–7) increases the negative charge on proteins.

problem	solution
Diffuse band patterns	Transfer immediately after electrophoretic separation. If equilibrating before the transfer, shorten or eliminate the equilibration time or move the gel to the cold room during equilibration.
	If transfer buffer contains methanol (≥10%), equilibrate the gel in transfer buffer for 30 minutes to allow it to shrink before assembling the stack. Note: Because methanol causes the gel to shrink slightly, large molecules may migrate more slowly.
	Take care that the gel is held firmly against the membrane and that it does not shift once contact is made.
	If excess heating occurs during the transfer, lower the temperature of the cooling fluid in the heat exchanger.
	Check that the preferred binding surface of the membrane (if any) contacts the gel.
Inefficient binding to membrane	
Chemical parameters	Fix or crosslink the molecule onto the membrane according to the requirements of the nucleic acid, protein, or membrane type.
	Prepare protein transfer buffer *without* SDS.
	Verify the optimal amount of methanol required for the membrane type and check the buffer solution. Add 10–20% methanol to the transfer buffer to enhance binding to nitrocellulose.
Membrane parameters	Wear gloves when handling membranes.
	Store membranes at ambient temperature out of direct sunlight to keep the membranes activated.
	Use a membrane with a smaller pore size (0.10–0.20 µm) if proteins pass through the membrane, or use a different membrane type.
	Place a membrane both over and under the gel if you suspect one protein is moving in the opposite direction from the majority of the proteins. Check both membranes for protein(s).
	Check if too much sample is available for the binding surface area by applying two membranes instead of one. If "blow through" occurs, reduce the sample load.

For more troubleshooting hints,
refer to Bjerrum, O.J. *et al.* (1988).

Electrotransfer notes

Electrophoretic transfer advantages

Electrophoretic transfer of proteins and nucleic acids is much faster than the blotting methods first described by Southern for DNA, Alwine *et al.* for RNA, or Renart *et al.* for proteins. The tank transfer method uses high current to reduce the transfer time of most samples to 45–60 minutes.

Electrophoretic transfer can improve transfer efficiency over non-electrophoretic blotting, especially for proteins, but no quantitative transfer technique has yet been developed due to the complexity of the reactions. Quantitative recovery is actually not required for most purposes because binding macromolecules to a membrane increases the sensitivity of detection methods such as autoradiography and permits detection of specific proteins by antibodies or affinity labels, and of specific nucleic acids by hybridization with complementary strands of RNA or DNA.

The buffer can be chosen to result in a transfer toward either the cathode or the anode. The buffer pH must be such that all species of interest are charged and migrate in the same direction. The ionic strength should not be too high, since this will produce excessive current and heat. For this reason, the high salt conditions used by Southern for capillary blotting of DNA cannot be used. The most widely used buffer systems are those of Towbin *et al.* for transferring proteins, and of Bittner *et al.* for transferring nucleic acids. Buffer systems for transfer of each type of sample are listed later in this section.

Factors affecting the transfer

Parameters such as sample characteristics, membrane type, gel pore size, and the transfer buffer used all contribute to the transferability of macromolecules, and should be kept in mind when developing a protocol. Very small molecular species, for instance, migrate quickly but often do not bind as well as larger molecules; large molecules bind more efficiently but do not elute from the gel as rapidly. The rate of elution is also affected by the pore size of the gel and the orientation of the molecules.

Further, the degree to which molecules bind to the membrane is influenced by membrane characteristics such as pore size and type, and buffer characteristics such as pH, salt type and concentration, and the presence of detergents such as sodium dodecyl sulfate (SDS). Conditions required for efficient elution may not coincide with optimal conditions for binding. To find the optimum conditions for transferring your sample, balance these effects: If the sample elution rate is slow, a longer transfer period may be required. (In our experience, low voltage transfers for longer periods do not offer much improvement.) If sample binding is inadequate, try different buffer conditions. For a comprehensive review, see Gershoni and Palade (1983).

If the transfer buffer system is different from the electrophoresis buffer system, the gel should be equilibrated with the transfer buffer before the transfer to ensure swelling or shrinking occurs before the gel contacts the transfer membrane. If this step is skipped, band distortion or loss of resolution could result.

Instrument guidelines

Cooling

Considerable Joule heat is generated during any transfer because of the high current employed, **so active cooling is recommended,** especially for transfers requiring more than one hour, protein transfers where biological activity must be retained, or transfer of nucleic acids. (The high conductivity of the phosphate buffer used by Bittner *et al.* (1980) leads to a relatively rapid temperature rise.) Buffer temperature should not exceed 45 °C because the cassettes and electrode supports may warp. Use a circulator bath set to 10 °C if using water as a coolant. (You can use a lower setting if the coolant is 50/50 ethylene glycol/water.) Never leave the unit unattended for more than one hour under high power conditions (>250 mA).

Power setting

If using a power supply that can be set to either constant current or constant voltage mode, we recommend that it be set to operate in constant current mode. Buffer conductivity increases with temperature. During blotting in an uncooled chamber, Joule heating and rising conductivity may result in dangerous overheating if the power supply is set to maintain constant voltage. If a constant voltage power supply must be used, monitor and adjust the voltage to maintain a current at or below 400 mA.

Protein transfers

Study summaries

Gershoni and Palade (1982) investigated factors affecting protein recovery from SDS gels to nitrocellulose or DBM paper. According to their findings, methanol in the Towbin buffer system is necessary to achieve efficient binding to nitrocellulose. Methanol improves binding in part by removing protein-bound SDS. In the absence of methanol, labeled bovine serum albumin (BSA) passes through at least five layers of membranes. Methanol may cause a gel to shrink, however, so the elution rate decreases. By using a cationic membrane (such as nylon), which binds the proteins more efficiently, and omitting methanol from the transfer buffer, Gershoni and Palade obtained a much more quantitative transfer. The disadvantage of cationic membrane is that protein stains also bind well, so that the staining background tends to be very high. Properly quenched, however, this paper can be used for antibody detection or other overlay methods of protein identification. A summary of membrane type and recommended methanol concentration follows:

membrane type	methanol %
Charged nylon	0
Nitrocellulose	≤ 20
PVDF	≤ 15

Some workers have reported to us that a low concentration of SDS (0.1%) improves the transfer of protein from an SDS gel. Burnette (1981) and Symington *et al.* (1981) investigated the effect of the molecular weight of protein. Gibson (1981) describes a method to increase the extent of transfer of large proteins by limited cleavage with pronase during transfer.

Protein transfer buffers

Use a buffer with low ionic strength, such as the two listed below, to prevent overheating. Use the alternate CAPS buffer when Tris cannot be used, as in peptide sequencing. CAPS can improve transfer because of its effect on the charge of the protein (see Matsudaira, 1987). For native proteins, we suggest using the electrophoresis buffer for transfer as well. Use the Towbin buffer to transfer SDS-denatured proteins toward the anode.

Towbin buffer

(25 mM Tris, 192 mM glycine, 20% v/v methanol, pH 8.3, 2 liters)

Tris (FW 121.1)	25 mM	6.0 g
Glycine (FW 75.07)	192 mM	28.8 g
SDS[a] (FW 288.4)	0.1% (3.5 mM)	2.0 g

Dissolve in 1.5 liters distilled water. Add methanol as required[b]. Bring to 2 liters with distilled water. Do not adjust the pH, which should be between 8.2 and 8.4. Optional: Chill before use.

[a]Optional: Adding SDS can improve transfer efficiency.

[b]Depending on the membrane type selected, adding methanol can improve the transfer results (see discussion and table above). Because buffers containing methanol may deteriorate if stored for long periods, add methanol as required just prior to transfer.

CAPS buffer, 1X

(10 mM CAPS, pH 11.0, 2 liters)

CAPS (FW 221.3) [3-(cyclohexylamino)-1-propanesulfonic acid]	10 mM	4.44 g

Dissolve in 1.5 liters distilled water, adjust to pH 11.0 with conc. NaOH. Adjust volume to 2.0 liters.

Nucleic acid transfers

Nucleic acids must normally be transferred in denatured form for most efficient binding. RNA is normally denatured with glyoxal before separation or separated in denaturing gels containing formaldehyde or methyl mercury. However, double stranded DNA is usually denatured in the gel with NaOH. The alkali must be neutralized and the gel equilibrated in transfer buffer before electrotransfer. For both DNA and RNA gels, any SDS must also be removed to assure efficient binding. Bittner *et al*. (1980) wash gels three times, 20 minutes each, to assure complete removal of denaturants and detergents.

See Bittner *et al.* for a study of the transfer efficiency for DNA of different sizes. The Bittner transfer buffer contains 25 mM sodium phosphate, pH 6.5. Also described is a method for the introduction of nicks by limited nuclease action in order to facilitate transfer of larger DNA fragments.

Recommended DNA buffers include the Bittner sodium phosphate buffer (see reference) and TBE. For RNA, TAE is recommended. TBE and TAE stock recipes are listed below. These buffers are most often diluted to 1X, but the concentration can range down to 0.1X. Cooling is strongly recommended for these buffers, especially at higher concentrations.

EDTA solution[a]

(0.5 M EDTA, pH 8.0, 100 ml)

$Na_2EDTA \cdot 2H_2O$ (FW 372.2)	0.5 M	18.6 g

Dissolve in 70 ml distilled water. Adjust to pH 8.0 with 10 M NaOH (approx. 5 ml), then add distilled water to 100 ml.

DNA transfer buffer, 10X

(10X Tris-borate-EDTA (TBE)[a], pH ~8.2, 1 liter)

Tris (FW 121.1)	900 mM	109.0 g
Boric acid (FW 61.83)	900 mM	55.6 g
EDTA solution (0.5 M, pH 8.0)	20 mM	40.0 ml

Distilled water to 1.0 liter. Do not adjust pH.

Dilute to 1X before use to yield 90 mM Tris, 90 mM boric acid, and 2 mM EDTA.

This dilution is commonly used, but dilutions down to 0.1X may be used should it be necessary to decrease the amount of current in the system in order to control overheating.

RNA transfer buffer, 10X

(10X Tris-acetate-EDTA (TAE)[b], pH ~8.4, 1 liter)

Tris (FW 121.1)	400 mM	48.4 g
Acetic acid, glacial (~17.4 M)	~200 mM	11.4 ml
EDTA solution (0.5 M, pH 8.0)	10 mM	20.0 ml

Distilled water to 1.0 liter. Do not adjust pH.

Dilute to 1X before use to yield 40 mM Tris, ~20 mM acetate, and 1 mM EDTA.
This dilution is commonly used, but dilutions down to 0.1X may be used should it be necessary to decrease the amount of current in the system in order to control overheating.

[a] *Current Protocols in Molecular Biology* (1993), A.2.1.

[b] Sambrook, J., and Russell, D.W. (2001) Molecular Cloning: A Laboratory Manual, A1.17.

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Ordering information

product	quantity	code no.
Hoefer TE 22 Tank Transfer Unit. Includes: 4 gel cassettes, 8 foam sponges, 3-mm thick, 4 foam sponges, 6-mm thick, 25 sheets of blotter paper	1	TE22
Accessories and replacement parts		
Gel cassette, 2 foam sponges, 3-mm thick, and 1 foam sponge, 6-mm thick	1	TE24
Foam sponges, 9 × 10.5 cm, 6-mm thick	4	TE25
Foam sponges, 9 × 10.5 cm, 3-mm thick	4	TE25F-1/8
Electrode panel	1	TE23
Safety lid with cables	1	TE29
High voltage leads with jacks	pair	SE6056-HV
Quick-fit coupler body, female, to fit 9.5 mm (3/8") ID tubing	2	QF3/8
Quick-fit coupler body, male, to fit 9.5 mm (3/8") ID tubing	2	QFX3/8
Blotter paper		
Blotter paper, sheets, 9 × 10.5 cm	50	TE26
Companion products		
Hoefer PS 2A200 Power Supply, 200 V, 2A	1	PS2A200

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